# **CFTR Genotype and Maximal Exercise Capacity in Cystic Fibrosis** A Cross-Sectional Study

Thomas Radtke<sup>1</sup>, Helge Hebestreit<sup>2</sup>, Sabina Gallati<sup>3</sup>, Jane E. Schneiderman<sup>4,5</sup>, Julia Braun<sup>1</sup>, Daniel Stevens<sup>6,7</sup>, Erik H. J. Hulzebos<sup>8</sup>, Tim Takken<sup>8</sup>, Steven R. Boas<sup>9</sup>, Don S. Urquhart<sup>10</sup>, Larry C. Lands<sup>11</sup>, Sergio Tejero<sup>12</sup>, Aleksandar Sovtic<sup>13</sup>, Tiffany Dwyer<sup>14,15</sup>, Milos Petrovic<sup>16</sup>, Ryan A. Harris<sup>17</sup>, Chantal Karila<sup>18</sup>, Daniela Savi<sup>19,20</sup>, Jakob Usemann<sup>21</sup>, Meir Mei-Zahav<sup>22</sup>, Elpis Hatziagorou<sup>23</sup>, Felix Ratjen<sup>4</sup>, and Susi Kriemler<sup>1</sup>; CFTR-Exercise Study Group

<sup>1</sup>Epidemiology, Biostatistics and Prevention Institute, University of Zurich, Zurich, Switzerland; <sup>2</sup>Paediatric Department, University Hospitals Würzburg, Würzburg, Germany; <sup>3</sup>Division of Human Genetics, Department of Pediatrics, Inselspital, University of Berne, Berne, Switzerland; <sup>4</sup>Division of Respiratory Medicine, Hospital for Sick Children, Toronto, Ontario, Canada; <sup>5</sup>Faculty of Kinesiology and Physical Education, University of Toronto, Toronto, Ontario, Canada; <sup>6</sup>Department of Pediatrics, Division of Respirology, Faculty of Medicine, Dalhousie University, Halifax, Nova Scotia, Canada; <sup>7</sup>School of Heatth and Human Performance, Faculty of Heatth, Dalhousie University, Halifax, Nova Scotia, Canada; <sup>7</sup>School of Medicine, Chicago, Illinois; <sup>10</sup>Department of Paediatric Respiratory and Sleep Medicine, Netherlands; <sup>9</sup>Northwestern University Feinberg School of Medicine, Chicago, Illinois; <sup>10</sup>Department of Paediatric Respiratory and Sleep Medicine, Royal Hospital for Sick Children, Edinburgh, United Kingdom; <sup>11</sup>Montreal Children's Hospital Virgen del Rocio, Sevilla, Spain; <sup>13</sup>Department of Pulmonology, Mother and Child Health Institute of Serbia, School of Medicine, University of Belgrade, Belgrade, Serbia; <sup>14</sup>Discipline of Physiotherapy, Faculty of Health Sciences, University of Sydney, Australia; <sup>15</sup>Department of Respiratory Medicine, Royal Prince Alfred Hospital, Vienna, Australia; <sup>16</sup>Derive Prevention Institute, Augusta University, Augusta, Georgia; <sup>18</sup>Service de pneumologie et allergologie pédiatriques, Hôpital Necker Enfants malades, Assistance Publique – Hôpitaux de Paris, University of Rome, Rome, Rome, taty; <sup>20</sup>Cystic Fibrosis Unit, Bambino Gesù Children's Hospital, Nenne, Italy; <sup>20</sup>Cystic Fibrosis Unit, Bambino Gesù Children's Hospital, Rome, Italy; <sup>21</sup>University Children's Hospital Basel, Basel, Switzerland; <sup>22</sup>Pulmonary Institute, Schneider Children's Medical Center, Sackler School of Medicine, Tel Aviv University, Tel Aviv, Israel; and <sup>23</sup>Paediatric Pulmonology and CF Unit, Paedia

ORCID ID: 0000-0002-1723-1070 (T.R.).

# Abstract

**Rationale:** Cystic fibrosis transmembrane conductance regulator (CFTR) is expressed in human skeletal muscle cells. Variations of CFTR dysfunction among patients with cystic fibrosis may be an important determinant of maximal exercise capacity in cystic fibrosis. Previous studies on the relationship between *CFTR* genotype and maximal exercise capacity are scarce and contradictory.

**Objectives:** This study was designed to explore factors influencing maximal exercise capacity, expressed as peak oxygen uptake  $(\dot{V}o_{2peak})$ , with a specific focus on *CFTR* genotype in children and adults with cystic fibrosis.

**Methods:** In an international, multicenter, cross-sectional study, we collected data on *CFTR* genotype and cardiopulmonary exercise tests in patients with cystic fibrosis who were ages 8 years and older. *CFTR* mutations were classified into functional classes I–V.

**Results:** The final analysis included 726 patients (45% females; age range, 8–61 yr; forced expiratory volume in 1 s, 16 to 123% predicted) from 17 cystic fibrosis centers in North America, Europe, Australia, and Asia, all of whom had both valid maximal cardiopulmonary

exercise tests and complete *CFTR* genotype data. Overall, patients exhibited exercise intolerance ( $\dot{Vo}_{2peak}$ , 77.3 ± 19.1% predicted), but values were comparable among different *CFTR* classes. We did not detect an association between *CFTR* genotype functional classes I–III and either  $\dot{Vo}_{2peak}$  (percent predicted) (adjusted  $\beta = -0.95$ ; 95% CI, -4.18 to 2.29; *P* = 0.57) or maximum work rate (Watt<sub>max</sub>) (adjusted  $\beta = -1.38$ ; 95% CI, -5.04 to 2.27; *P* = 0.46) compared with classes IV–V. Those with at least one copy of a *F508del-CFTR* mutation and one copy of a class V mutation had a significantly lower  $\dot{Vo}_{2peak}$  ( $\beta = -8.24\%$ ; 95% CI, -14.53 to -2.99; *P* = 0.003) and lower Watt<sub>max</sub> (adjusted  $\beta = -7.59\%$ ; 95% CI, -14.21 to -0.95; *P* = 0.025) than those with two copies of a class II mutation. On the basis of linear regression analysis adjusted for relevant confounders, lung function and body mass index were associated with  $\dot{Vo}_{2peak}$ .

**Conclusions:** *CFTR* functional genotype class was not associated with maximal exercise capacity in patients with cystic fibrosis overall, but those with at least one copy of a *F508del-CFTR* mutation and a single class V mutation had lower maximal exercise capacity.

**Keywords:** cystic fibrosis transmembrane conductance regulator; peak oxygen uptake; lung disease; cardiorespiratory fitness

(Received in original form July 18, 2017; accepted in final form November 15, 2017)

Members of the CFTR-Exercise Study Group can be found before the REFERENCES.

This article has an online supplement, which is accessible from this issue's table of contents at www.atsjournals.org.

Ann Am Thorac Soc Vol 15, No 2, pp 209–216, Feb 2018 Copyright © 2018 by the American Thoracic Society DOI: 10.1513/AnnalsATS.201707-570OC Internet address: www.atsjournals.org

Cystic fibrosis (CF) is a disorder with autosomal recessive inheritance that ultimately leads to respiratory failure and premature death. The disease is caused by mutations in the cystic fibrosis transmembrane conductance regulator (CFTR) gene coding for the CFTR protein that functions as an ion channel, mainly for chloride and bicarbonate (1). CFTR mutations are grouped into different classes based on their effect on CFTR protein production, trafficking, function, and stability (1). CFTR is expressed in cell membranes of epithelial cells. It can also be found at the level of the sarcoplasmic reticulum in airway smooth muscle (2), human skeletal muscle cells (3, 4), and myocardium (5), but the precise physiological role of CFTR in these tissues is not fully understood (6). Previous studies suggest intrinsic functional abnormalities in skeletal muscles lacking functional CFTR (3, 4), possibly based on a disturbed calcium homeostasis in the muscle and increased systemic inflammation (3). It is therefore plausible that dysfunctional CFTR in human skeletal muscle could be a factor contributing to peripheral muscle weakness (7, 8) and reduced exercise capacity (e.g., peak oxygen uptake [VO<sub>2peak</sub>]) in CF (8, 9).

Because VO<sub>2peak</sub> is a predictor of mortality in CF (10), knowledge of important correlates of aerobic performance may help to guide patient care. Few, predominantly small, studies have previously been done to investigate the relationships between CFTR genotype and Vo<sub>2peak</sub> in patients with CF (11-14), with equivocal and controversial results. Selvadurai and colleagues (12) found that children with CF carrying an F508del-CFTR gene mutation on one allele and a class I or class II CFTR mutation on the second allele had lower VO<sub>2peak</sub> (approximately 30 to 45%) and peak anaerobic exercise capacity (approximately 10 to 17%) than children with a class III, IV, or V mutation on the second CFTR allele. However, this finding was based on univariate analysis of a relatively small sample size and thus did not control for relevant determinants

of exercise capacity, such as nutritional status and chronic infection with *Pseudomonas aeruginosa* (8, 13, 15), which could have influenced their study findings and interpretation.

The present study was designed to investigate factors associated with  $\dot{\rm Vo}_{2\rm peak}$ (primary endpoint) and maximum work rate (Watt<sub>max</sub>; secondary endpoint) with a specific focus on *CFTR* genotype in a large international cohort of children and adults with CF. We aimed to compare maximal exercise capacity among patients with *CFTR* mutations that result in severely reduced function (combinations of classes I–III mutations) with that of patients with combinations of classes IV–V mutations characterized by some residual CFTR function at the cell surface (16).

# Methods

## **Study Design and Patients**

We invited members from the Exercise Working Group of the European Cystic Fibrosis Society to participate in the study. We also searched the PubMed database for publications on exercise testing in CF to identify clinical studies that included cardiopulmonary exercise testing (CPET). The CPET had to be performed on a cycle ergometer employing the Godfrey cycle protocol (17) or a modification thereof. Inclusion and exclusion criteria and detailed information on CPET can be found in the online supplement.

We collected data on anthropometric characteristics, CF-related comorbidities (e.g., exocrine pancreatic insufficiency, cystic fibrosis-related diabetes [CFRD], colonization with *P. aeruginosa*), pulmonary function, CPET-related data, and genetic data. Chronic *P. aeruginosa* infection was considered to be present when more than 50% of at least four sputum samples collected in the previous year were positive (18). Spirometry was performed according to American Thoracic Society/European Respiratory Society standards (19). We calculated percent predicted values for spirometry (20),  $\dot{V}_{0_{2peak}}$  (21, 22), Watt<sub>max</sub> (21, 23), and peak heart rate (24), as well as *z*-scores for body mass index (BMI) based on World Health Organization criteria (25). Percent body fat was calculated using sex- and agespecific prediction equations (26), and lean body mass was derived from body fat and weight. For the final models,  $\dot{V}_{0_{2peak}}$ (primary endpoint) and Watt<sub>max</sub> (secondary endpoint) were expressed as percent predicted of reference values (22, 23).

The classification of CFTR genotype (27, 28) was performed by a geneticist (S.G.) who was blinded to the exercise testing data. Details about the functional classification of CFTR alleles are shown in Table E1 in the online supplement. Study participants were grouped for the analysis of the primary endpoint (Vo<sub>2peak</sub> percent predicted) in patients with both CFTR alleles in either class I, II, or III (corresponding to severely reduced CFTR function) and patients with at least one mutant allele in class IV or class V (corresponding to some residual CFTR function) (16). In an exploratory analysis, we also compared maximal exercise capacity between patients with at least one copy of the F508del-CFTR mutation and categorized them into five groups on the basis of their second CFTR mutation class (11, 27).

Ethical approval for this study was obtained from the cantonal ethical committee of Zurich, Switzerland (2015-0109). All centers obtained ethical approval (if required) for the use of their anonymized patient data according to national and local policies.

#### **Statistical Analysis**

Details on the statistical analyses can be found in the online supplement. We used analysis of variance, Kruskal–Wallis tests, and  $\chi^2$  tests to compare variables between groups. We used mixed-effects models with a random intercept for study center to identify characteristics associated with  $\dot{V}O_{2peak}$  and Watt<sub>max</sub> (percent predicted). Multilevel mixed-effects models with a random intercept for each study center were used to examine associations between *CFTR* genotype-based group and

Author Contributions: Conception and design: T.R., S.K., and H.H.; acquisition of data: A.S., C.K., D. Stevens, D. Savi, D.S.U., E.H., E.H.J.H., F.R., H.H., J.E.S., L.C.L., M.M.-Z., M.P., R.A.H., S.K., S.R.B., T.D., S.T., and T.T.; genotype classification: S.G. and T.R.; statistical analysis: J.B. and T.R.; interpretation: T.R., H.H., F.R., J.B., J.U., and S.K.; and first draft: T.R., H.H., and S.K. All authors edited, reviewed, and approved the final version of the manuscript.

Correspondence and requests for reprints should be addressed to Thomas Radtke, Ph.D., Epidemiology, Biostatistics and Prevention Institute (EBPI), University of Zurich, Hirschengraben 84, 8001 Zurich, Switzerland. E-mail: thomas.radtke@uzh.ch.

VO<sub>2peak</sub> and Watt<sub>max</sub> (percent predicted). The models were a priori adjusted for the variables age, sex, BMI z-score, and forced expiratory volume in 1 second (FEV<sub>1</sub>; percent predicted) because it is generally accepted that they have an influence on  $\dot{V}O_{2peak}$  (9, 29, 30). According to the results reported by van de Weert-van Leeuwen and colleagues (13), chronic infection with P. aeruginosa is associated with decline in  $\dot{V}O_{2peak}$  over time and therefore represents an important covariate in our model. To decide if the potential confounders CFRD or pancreatic insufficiency should be taken into account, the Akaike information criterion was used for model choice. However, the models did not improve when we added those variables, for which reason they were discarded. We also checked if interactions between CFTR genotypebased group and the other influential variables improved the models, which was not the case. To confirm our approach of defining the exercise capacity outcome variables as percent predicted values, we additionally performed linear mixed models using a multiplicative, allometric approach to account for potential effects of body size and pulmonary function on the relationship between CFTR genotype and VO<sub>2peak</sub> (see online supplement).

# Results

We contacted 32 centers in North America, Europe, Australia, and Asia by e-mail, 17 of which provided data for this study. Reasons for nonparticipation can be found in the online supplement. Included centers were in Canada (n = 3; 293 patients); the United States (n = 2; 110 patients); and one center each from Austria (n = 30)patients), Australia (n = 30 patients), France (n = 59 patients), Germany (n = 69patients), Greece (n = 39 patients), Israel (n = 37 patients), Italy (n = 34 patients), the Netherlands (n = 93 patients), Spain (n = 51 patients), Serbia (n = 64 patients), Switzerland (n = 42 patients), and the United Kingdom (n = 39 patients). A flowchart of included patients is shown in Figure 1, and patient characteristics are shown in Table 1. Of 838 patients, 112 did not reach their maximal exercise level during the CPET and were excluded from further analyses. Criteria for a maximal



Figure 1. Flowchart of included patients. CFTR = cystic fibrosis transmembrane conductance regulator; CPET = cardiopulmonary exercise test.

effort during CPET are provided in the online supplement. These 112 patients were, on average, older, had lower FEV<sub>1</sub>, lower exercise capacity, and more CF-related comorbidities (Table 1). The lower exercise capacity in patients not performing maximally was not explained by a pulmonary function limitation (i.e., a lower breathing reserve at end exercise [VEpeak/predicted maximum voluntary ventilation] than the group demonstrating maximal effort) (Table 1). The final dataset included 726 patients (45% females; age,  $18.7 \pm 8.5$  yr; age range, 8–61 yr; FEV<sub>1</sub>, 76.6  $\pm$  22.9; FEV<sub>1</sub> range, 16–123; Vo<sub>2peak</sub>, 77.3  $\pm$  19.1% predicted;  $\dot{V}O_{2peak}$  range, 25-137). Figure 1 in the online supplement shows the relationship between FEV<sub>1</sub> and VO<sub>2peak</sub>. Table 2 shows data for maximal exercise capacity, lung function, nutritional status, and CPET-related variables grouped by CFTR mutation class.

#### Predictors of Maximal Exercise Capacity in Patients with Cystic Fibrosis

In the univariate analysis adjusted for study center, we did not find any association of *CFTR* functional class (I–III vs. IV–V) with either  $\dot{V}o_{2peak}$  percent predicted ( $\beta = 2.35$ ; 95% CI, -1.27 to 6.35; P = 0.19) or Watt<sub>max</sub> percent predicted ( $\beta = 2.34$ ; 95% CI, -1.90

to 6.58; P = 0.28). In contrast, age, FEV<sub>1</sub> percent predicted, BMI z-score, CFRD, and chronic P. aeruginosa infection were associated with VO<sub>2peak</sub> and Watt<sub>max</sub> (expressed as percent predicted). Exocrine pancreatic insufficiency was associated only with Wattmax (percent predicted). Forced vital capacity was not included in the final multilevel multivariate model, owing to a high collinearity with FEV<sub>1</sub> (R = 0.89; P = 0.001). We noted differences in  $\dot{V}O_{2peak}$ and Watt<sub>max</sub> among study centers. Consequently, center was included as a random intercept in the multilevel mixed-effects models. In fully adjusted models, FEV<sub>1</sub> percent predicted ( $\beta = 0.41$ ; 95% CI, 0.36 to 0.47; P < 0.001) and BMI *z*-score ( $\beta = 1.75$ ; 95% CI, 0.76 to 2.74; P = 0.001) were significantly associated with VO<sub>2peak</sub> (percent predicted). In the model for Watt<sub>max</sub> (percent predicted), FEV<sub>1</sub> percent predicted ( $\beta = 0.46$ ; 95% CI, 0.40–0.52; *P* < 0.001) and BMI *z*-score  $(\beta = 1.27; 95\% \text{ CI}, 0.15-2.38; P = 0.03)$ remained the only significant predictors.

## Associations of CFTR Functional Class and Maximal Exercise Capacity

The prevalence of CFRD, pancreatic insufficiency, and *P. aeruginosa* infection were higher for patients with only classes I–III *CFTR* mutations than for patients

 Table 1. Clinical characteristics and cardiopulmonary exercise testing data for patients with valid versus nonvalid maximal exercise tests

Variable	n	Valid Maximal CPET	n	Nonvalid Maximal CPET
Age, yr* Sex, % females Body mass index, z-score <sup>†</sup> FEV <sub>1</sub> , % predicted <sup>†</sup> Cystic fibrosis–related diabetes, $n$ (%) <sup>†</sup> <i>Pseudomonas aeruginosa</i> , $n$ (%)* Pancreatic insufficiency, $n$ (%) Vo <sub>2peak</sub> , % predicted* (22) Vo <sub>2peak</sub> , % predicted* (21, 22) Watt <sub>max</sub> , % predicted* (23) Watt <sub>max</sub> , % predicted* (21, 23) HR <sub>max</sub> , % predicted* Respiratory exchange ratio* VE <sub>peak</sub> /MVV <sub>pred</sub> *	726 726 724 723 716 724 726 726 724 725 $639^{\ddagger}$ 725	$\begin{array}{c} 16.4 \ (13.0,\ 22.1) \\ 330 \ (45) \\ -0.24 \ (-0.92,\ 0.47) \\ 79 \ (66,\ 91) \\ 79 \ (11) \\ 371 \ (52) \\ 611 \ (84) \\ 79 \ \pm \ 19 \\ 80 \ \pm \ 20 \\ 85 \ \pm \ 22 \\ 91 \ \pm \ 23 \\ 93 \ (88,\ 96) \\ 1.16 \ (1.10,\ 1.23) \\ 86 \ (70,\ 102) \end{array}$	112 112 112 111 109 105 112 112 112 112 112 112 112 112 36 <sup>‡</sup> 111	$\begin{array}{c} 20.9 \ (13.9, \ 28.8) \\ 45 \ (40) \\ -0.01 \ (-0.75, \ 0.70) \\ 74 \ (56, \ 86) \\ 20 \ (18) \\ 83 \ (79) \\ 90 \ (80) \\ 67 \pm 15 \\ 70 \pm 15 \\ 74 \pm 18 \\ 82 \pm 19 \\ 87 \ (80, \ 92) \\ 0.99 \ (0.96, \ 1.01) \\ 76 \ (63, \ 87) \end{array}$

Definition of abbreviations: CPET = cardiopulmonary exercise test;  $FEV_1$  = forced expiratory volume in 1 second;  $HR_{max}$  = maximum heart rate;  $V_{E_{peak}}$  = peak minute ventilation;  $V_{o_{2peak}}$  = peak oxygen consumption;  $Watt_{max}$  = maximum work rate.

Data are mean  $\pm$  SD, median (interquartile range), or number (percent). Percent predicted values for Vo<sub>2peak</sub> were calculated using equations from Orenstein (22) and Jones and colleagues (21). Data for Vo<sub>2peak</sub> are shown separately according to Orenstein (22) and a combination of Orenstein (22) and Jones and colleagues (21) for patients aged 16 years or younger and patients older than 16 years of age, respectively. Watt<sub>max</sub> percent predicted values were calculated using equations from Godfrey and colleagues (23) and Jones and colleagues (21). Data are shown separately according to Godfrey and colleagues (23) and Jones and colleagues (21). Data are shown separately according to Godfrey and colleagues (23) and Jones and colleagues (21) for patients aged 16 years or younger and patients older than 16 years of age, respectively. Statistical comparisons between groups were performed using an independent *t* test, a nonparametric Mann–Whitney *U* test, or the  $\chi^2$  test as appropriate.

\**P* < 0.001. †*P* < 0.05.

<sup>‡</sup>Respiratory exchange ratio data from two study centers were not available.

with at least one CFTR allele associated with residual function (class IV or class V). Univariate analysis revealed no differences in maximal exercise capacity (absolute and percent predicted Vo<sub>2peak</sub> and Watt<sub>max</sub>) between the two groups (Table 3). In mixed-effects models, we did not detect an association between CFTR genotype functional class and either  $\dot{V}O_{2\text{peak}}$  (percent predicted) ( $\beta = -0.95$ ; 95% CI, -4.18 to 2.29; P = 0.57) or Watt<sub>max</sub> ( $\beta = -1.38$ ; 95% CI, -5.04 to 2.27; P = 0.46). In these models, both lung function and nutritional status were associated with VO<sub>2peak</sub> and Wattmax (percent predicted) independent of CFTR function (Table 4). These results were confirmed when allometric modeling was applied (Table E2) and when exercise capacity was expressed per kilogram of body weight or lean body mass (data not shown). Moreover, the results were similar when the 112 patients with nonmaximal CPETs were included in the analysis. In 835 and 831 patients, we found no association between CFTR genotype functional class and either  $\dot{V}\mathrm{O}_{2peak}$  (percent predicted)  $(\beta = -1.95; 95\%$  CI, -5.28 to 1.39; P = 0.25)or  $Watt_{max}$  ( $\beta = -2.42$ ; 95% CI, -5.97 to 1.48; *P* = 0.24), respectively.

In a subanalysis restricted to patients with preserved lung function (FEV<sub>1</sub>,  $\geq$ 80% predicted) and nutritional status (BMI *z*-score,  $\geq$ 50th percentile), we found no between-group differences in maximal exercise capacity in either univariate or adjusted analysis (Tables E3 and E4). Furthermore, a subanalysis on patients with moderate to severe lung function impairment (FEV<sub>1</sub>,  $\leq$ 60% predicted) did not show between-group differences in either  $\dot{V}o_{2peak}$  or Watt<sub>max</sub> (percent predicted) (Tables E5 and E6).

# Exercise Capacity among Patients with at Least One Copy of the *F508del-CFTR* Mutation

In 653 patients who had at least one copy of the *F508del-CFTR* (class II) mutation grouped in five classes according to their second allele, no differences among *CFTR* genotype and either  $\dot{\rm Vo}_{2peak}$  or Watt<sub>max</sub> were observed (absolute values or % predicted) (*see* Table E7). In mixed models, patients with one copy of a class V mutation had a significantly lower  $\dot{\rm Vo}_{2peak}$  than patients with two copies of a class II mutation ( $\beta = -8.24\%$ ; 95% CI, -14.53 to -2.99; P = 0.003) (*see* Table E8).

Furthermore, Watt<sub>max</sub> values were lower for the group with one copy of a class V mutation ( $\beta = -7.59\%$ ; 95% CI, -14.21 to -0.95; P = 0.025). In addition, the same association was found for  $\dot{V}o_{2peak}$  and Watt<sub>max</sub> (31) using allometric models (Table E9).

# Discussion

In this largest international multicenter study of exercise testing in patients with CF to date, we focused primarily on the relationship between *CFTR* genotype and predictors of maximal exercise capacity in patients with CF. We found that severity of *CFTR* genotype, using different CFTR categorizations, combinations, and analytic approaches, was not associated with maximal exercise capacity. In contrast, pulmonary function and nutritional status, expressed as BMI, were strongly associated with exercise capacity.

Our data do not support a relevant role of *CFTR* genotype on maximal exercise capacity in patients with CF. However, some theoretical considerations deserve further explanation to shed light on **Table 2.** Lung function, nutritional status, and exercise capacity in 726 patients according to *CFTR* mutation (functional classes I–V), based on functional defect of the milder of the two mutations

	CFTR Class I/I	CFTR Class ≤II/II	CFTR Class ≤III/III	CFTR Class ≤IV/IV	CFTR Class ≤V/V
No. of patients	32	550	39	63	42
Age, yr*	14.6 (12.7, 18.3)	16.2 (12.9, 21.3)	16.7 (12.1, 25.0)	16.6 (12.1, 25.0)	19.0 (14.8, 26.5)
Sex, % female	14 (44)	244 (44)	16 (41)	34 (54)	21 (50)
diabetes, n (%)*	7 (22)	64 (12)	4 (10)	2 (3)	1 (2)
Pancreatic insufficiency, $n (\%)^{\dagger}$	35 (97)	529 (93)	34 (89)	15 (24)	10 (24)
Pseudomonas aeruginosa, n (%)*	32 (100)	519 (95)	21 (55)	23 (37)	15 (36)
Body mass index, <sup>†</sup> kg/m <sup>2</sup>	18.8 (16.9, 20.1)	19.3 (17.3, 21.5)	20.4 (17.5, 24.2)	20.6 (18.8, 23.0)	22.3 (19.2, 25.0)
Body mass index, z-score*	-0.35 (-1.05, 0.39)	-0.28 (-0.97, 0.35)	0.26 (-0.53, 1.04)	-0.20 (-0.76, 0.63)	0.08 (-0.79, 1.14)
Lean body mass, kg*	38.2 (30.0, 46.4)	41.3 (32.9, 50.3)	43.5 (32.1, 55.2)	42.3 (36.7, 52.8)	47.0 (40.4, 61.1)
Body fat, % <sup>†</sup>	$17.2 \pm 4.7$	$18.2 \pm 5.7$	$19.9 \pm 5.5$	$21.4 \pm 6.4$	$22.4 \pm 6.5$
FEV <sub>1</sub> , % predicted	80 (45, 93)	79 (60, 94)	78 (50, 90)	86 (72, 96)	80 (62, 94)
Vo <sub>2peak</sub> , L/min	1.6 (1.3, 1.8)	1.7 (1.4, 2.3)	1.8 (1.3, 2.2)	1.8 (1.5, 2.3)	1.7 (1.3, 2.4)
Vo <sub>2peak</sub> , % predicted (22)	74 ± 17	$79 \pm 19$	$78 \pm 24$	83 ± 18	$74 \pm 19$
Vo <sub>2peak</sub> , % predicted (21, 22)	74 ± 17	80 ± 19	$79 \pm 23$	85 ± 18	$78 \pm 23$
$Vo_{2peak}$ , <82% predicted, <i>n</i> (%)	21 (66)	313 (57)	23 (59)	32 (51)	28 (67)
Watt <sub>max</sub> , W	111 (83, 140)	127 (98, 170)	130 (95, 163)	124 (95, 170)	130 (85, 180)
Watt <sub>max</sub> , % predicted (23)	77 ± 17	86 ± 22	$85\pm25$	86 ± 21	$78\pm20$
Watt <sub>max</sub> , % predicted* (21, 23)	79 ± 18	$92 \pm 24$	91 ± 27	92 ± 22	$87\pm20$
Watt <sub>max</sub> , $<$ 93% predicted, <i>n</i> (%)	27 (84)	358 (65)	23 (59)	44 (70)	31 (74)
HR <sub>max</sub> , % predicted	92 (86, 96)	93 (88, 97)	91 (87, 97)	92 (87, 96)	92 (86, 96)
Respiratory exchange ratio	1.18 (1.12, 1.25)	1.16 (1.10, 1.23)	1.16 (1.11, 1.24)	1.14 (1.09, 1.20)	1.17 (1.08, 1.23)
VE <sub>peak</sub> /MVV <sub>pred</sub> , %*	81 (72, 106)	88 (71, 103)	95 (72, 118)	75 (64, 92)	80 (61, 101)

Definition of abbreviations: CFTR = cystic fibrosis transmembrane conductance regulator;  $FEV_1$  = forced expiratory volume in 1 second;  $HR_{max}$  = maximum heart rate;  $MVv_{pred}$  = predicted maximum voluntary ventilation (calculated as  $FEV_1 \times 35$ );  $Ve_{peak}$  = peak minute ventilation;  $Vo_{2peak}$  = peak oxygen consumption;  $Watt_{max}$  = maximum work rate.

Data are mean  $\pm$  SD, median (interquartile range), or number (percent). Percent predicted values for  $\dot{V}o_{2peak}$  were calculated using equations from Orenstein (22) and Jones and colleagues (21). Data for  $\dot{V}o_{2peak}$  are shown separately according to Orenstein (22) and a combination of Orenstein (22) and Jones and colleagues (21) for patients aged 16 years or younger and patients older than 16 years of age, respectively. Watt<sub>max</sub> percent predicted values were calculated using equations from Godfrey and colleagues (23) and Jones and colleagues (21). Data are shown separately according to Godfrey and colleagues (23) and Jones and colleagues (21). Data are shown separately according to Godfrey and colleagues (23) and a combination of Godfrey and colleagues (23) and Jones and colleagues (21). Data are shown separately according to Godfrey and colleagues (23) and Jones and colleagues (21) for patients aged 16 years or younger and patients older than 16 years of age, respectively. Reduced exercise capacity ( $\dot{V}o_{2peak} < 82\%$  predicted and Watt<sub>max</sub> <93\% predicted) was calculated according to Nixon and colleagues (10). Statistical comparisons between different CFTR classes were performed using analysis of variance, a nonparametric Kruskal–Wallis test, or the  $\chi^2$  test for categorical variables.

\*P < 0.05, significantly different between CFTR classes.

 $^{+}P < 0.001$ , significantly different between CFTR classes.

potential underlying molecular mechanisms of the CFTR defect and its potential consequences on peripheral muscle function and exercise capacity. Recently, the expression of functional CFTR channels in human skeletal muscle was demonstrated (3, 4). Lamhonwah and colleagues (4) speculated that a dysfunction in sarcoplasmic reticulum CFTR channels might affect calcium release and thus impact the adenosine triphosphatemediated actin-myosin interaction that is essential for muscle contraction. Moreover, Divangahi and colleagues (3) suggested that excessive systemic inflammation initiates a process in which the abnormal vulnerability of CFTRdeficient muscle to proinflammatory mediators could play a key role in the development of skeletal muscle weakness

observed in individuals with CF (7). Of note, these observations are based on animal experiments and have not yet been confirmed in humans. On the basis of the in vitro experiments suggesting that systemic inflammation in combination with CFTR dysfunction may impair skeletal muscle function, it is reasonable to speculate that potential detrimental effects of CFTR genotype on maximal exercise capacity would be detected only with high levels of inflammation. However, although the present study included patients with mildly impaired pulmonary function (mean  $\pm$  SD FEV<sub>1</sub>, 77.3  $\pm$  23% predicted), a subgroup analysis including only patients with moderate to severe lung disease (FEV<sub>1</sub>, ≤60% predicted) yielded similar results for the comparison between minimal function and residual function mutations (Tables E5

and E6). However, this subanalysis is limited by the small sample size of patients with residual function mutations and should be interpreted with caution. It remains to be shown in larger (ideally prospective) studies whether our findings are applicable to patients with more advanced disease and substantial inflammation.

Our main aim was to compare maximal exercise capacity between patients with CF carrying only minimal function *CFTR* mutations (classes I–III) and those with at least one residual function (class IV or V) *CFTR* mutation. In the primary analysis, our data showed no differences in maximal exercise capacity between groups in both unadjusted and adjusted analyses. These results were confirmed by the use of allometric models that were computed to exclude potential limitations of the prediction **Table 3.** Clinical characteristics and cardiopulmonary exercise testing data between patients with two *CFTR* mutations in class I, II, or III compared with patients with at least one mutation in class IV or V

No. of patients         621         105           CFTR modulator therapy, n (%)         9 (1)         3 (3)           Age, yr         16.2 (12.9, 21.6)         18.0 (13.0, 25)	Variable	CFTR CFTR Classes I–III Classes IV–V
Sex, % female273 (44)55 (52)Cystic fibrosis-related diabetes, $n$ (%)*75 (12)3 (3)Pancreatic insufficiency, $n$ (%)*585 (95)25 (24)Pseudomonas aeruginosa, $n$ (%)*332 (54)38 (36)Body mass index, z-score* $-0.25$ ( $-0.95$ , $0.42$ ) $-0.11$ ( $-0.77$ ,Body fat, %*18.2 ± 5.721.8 ± 6.4Lean body mass, kg41.1 (32.5, 50.3)44.1 (36.4, 54)FEV1, % predicted79 (59, 93)84 (68, 96)Vo <sub>2peak</sub> , L/min1.74 (1.4, 2.2)1.78 (1.4, 2.4)Vo <sub>2peak</sub> , % predicted (21, 22)80 ± 1982 ± 20Wattmax, W125 (95, 168)130 (94, 176)Wattmax, % predicted (21, 23)91 ± 2490 ± 21HRmax, % predicted (21, 23)93 (88, 96)92 (87, 96)Respiratory exchange ratio1.16 (1.10, 1.23)1.15 (1.09, 1.Venext/MVVnred, %*88 (71, 104)78 (63, 96)	No. of patients DFTR modulator therapy, $n$ (%) Age, yr Sex, % female Dystic fibrosis-related diabetes, $n$ (%)* Pancreatic insufficiency, $n$ (%)* Pseudomonas aeruginosa, $n$ (%)* Dody mass index, z-score* Body fat, % <sup>†</sup> Lean body mass, kg FEV <sub>1</sub> , % predicted $V_{02peak}$ , % predicted (22) $V_{02peak}$ , % predicted (21, 22) Watt <sub>max</sub> , % predicted (23) Watt <sub>max</sub> , % predicted (21, 23) HR <sub>max</sub> , % predicted Respiratory exchange ratio $V_{Ereak}/MVV_{pred}$ , %*	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

*Definition of abbreviations*: CFTR = cystic fibrosis transmembrane conductance regulator; FEV<sub>1</sub> = forced expiratory volume in 1 second; HR<sub>max</sub> = maximum heart rate; MVV<sub>pred</sub> = predicted maximum voluntary ventilation (calculated as FEV<sub>1</sub> × 35);  $\dot{V}_{E_{peak}}$  = peak minute ventilation;  $\dot{V}_{O_{2peak}}$  = peak oxygen consumption; Watt<sub>max</sub> = maximum work rate.

Data are mean  $\pm$  SD, median (interquartile range), or number (percent). Percent predicted values for  $Vo_{zpeak}$  were calculated using equations from Orenstein (22) and Jones and colleagues (21). Data for  $Vo_{zpeak}$  are shown separately according to Orenstein (22) and a combination of Orenstein (22) and Jones and colleagues (21) for patients aged 16 years or younger and patients older than 16 years of age, respectively. Watt<sub>max</sub> percent predicted values were calculated using equations from Godfrey and colleagues (23) and Jones and colleagues (21). Data are shown separately according to Godfrey and colleagues (23) and Jones and colleagues (21). Data are shown separately according to Godfrey and colleagues (23) and a combination of Godfrey and colleagues (23) and Jones and colleagues (21). Data are shown separately according to Godfrey and colleagues (23) and a combination of Godfrey and colleagues (23) and Jones and colleagues (21). Data are shown separately according to Godfrey and colleagues (23) and a combination of Godfrey and colleagues (23) and Jones and colleagues (21). Data are shown separately according to Godfrey and colleagues (23) and a combination of Godfrey and colleagues (23) and Jones and colleagues (21) for patients older than 16 years of age, respectively. Statistical comparisons between the two groups were performed using an independent *t* test, a nonparametric Mann-Whitney *U* test, or the  $\chi^2$  test, as appropriate.

\*P < 0.05, significantly different between CFTR classes.

 $^{\dagger}P < 0.001$ , significantly different between CFTR classes.

equations for  $\dot{V}o_{2peak}$  and  $Watt_{max}$  (32). Regardless of the analytic approach, including adjustments for important confounders, we found robust evidence that *CFTR* genotype severity was not related to impaired maximal exercise capacity in patients with CF.

Our findings support previous studies showing that pulmonary function limitation and inadequate nutrition contribute to exercise intolerance in patients with CF (9, 13, 29, 30, 33, 34). Nutritional status plays a critical role in the progression of lung disease. Compared with healthy subjects, patients with CF exhibit peripheral muscle weakness (35) that is associated with reduced aerobic exercise capacity (7, 8). Thus, adequate nutrition and maintenance of BMI, and in particular muscle mass (35), are important for preserving exercise tolerance in patients with CF. The strong impact of lung function and nutritional status could mask the effect of *CFTR* genotype on exercise capacity in patients with CF. Consequently, to exclusively study the role of *CFTR* genotype on exercise capacity, we performed a subanalysis restricted to patients with preserved lung function (FEV<sub>1</sub>,  $\geq$ 80% predicted) and nutritional status (BMI *z*-score,  $\geq$ 50th percentile), and the results remained qualitatively the same.

Our data are in contrast to a study by Selvadurai and colleagues (12), who reported a relationship between the severity of CFTR functional impairment and reduced exercise capacity using univariate analysis in children with CF aged 8–17 years with at least one copy of the *F508del*-*CFTR* mutation. In univariate analysis, we noticed no differences between groups in either absolute or percent predicted values for  $\dot{V}O_{2peak}$  or  $Watt_{max}$  (Table E6). However, in the adjusted analysis, patients with one copy of a class V mutation had significantly lower exercise capacity (about 8% predicted for both VO<sub>2peak</sub> and Watt<sub>max</sub>) than patients homozygous for the F508del-CFTR mutation (Tables E7 and E8). Patients with one copy of a class V mutation did not differ in ethnicity and lung function but had better nutritional status compared with patients homozygous for the F508del-CFTR mutation. However, there was no evidence for ventilatory limitation (i.e., lower breathing reserve) during exercise in these patients, which may suggest that the lower exercise performance was reflective more of fitness level than of pulmonary status. Moreover, in mixed models, the presence of P. aeruginosa infection was significantly associated with a lower VO<sub>2peak</sub> and Watt<sub>max</sub> (Table E8). This is supported by a study by van de Weert-van Leeuwen and colleagues (13), who have previously shown that the presence of P. aeruginosa in adolescents with CF is a predictor of a steeper decline in VO<sub>2peak</sub> over time, independent of age, nutritional status, pulmonary function, and immunoglobulin G levels. Nevertheless, relatively fewer patients with a group V mutation were colonized with P. aeruginosa than patients with solely CFTR class II mutations (reference group), despite a lower VO<sub>2peak</sub> in the former than in the latter group. The differences between our study and the study by Selvadurai and colleagues (12) might be explained by the preserved lung function in the present cohort of patients and our rigorous adjustment for important clinical confounders in the statistical models. In addition, substantial CFTR genotype misclassification affecting different groups in the study by Selvadurai and colleagues (12) may contribute to the diverse findings. Finally, we can only speculate that lower habitual physical activity, a determinant of  $\dot{V}o_{2peak}$  in CF (9), contributes to the reduced exercise capacity in patients with one copy of a class V mutation. The cohort of patients with a class IV or V CFTR mutation in our study might be a subgroup of patients that have been diagnosed at older ages (i.e., when their CF disease progressed and started to become symptomatic). Thus, later CF diagnosis and less focus clinically on exercise and physical activity may affect

**Table 4.** Mixed models for association between patients with two *CFTR* mutations in class I, II, or III (group I–III) compared with patients with at least one mutation in class IV or V (group IV–V)

	$\beta$ -Coefficient (95% CI)	SE	P Value
Vo <sub>2peak</sub> , % predicted Age Sex <i>Pseudomonas aeruginosa</i> Body mass index, <i>z</i> -score FEV <sub>1</sub> , % predicted CFTR group	-0.14 (-0.32 to 0.04) -1.29 (-3.46 to 0.88) -1.93 (-4.44 to 0.58) 1.78 (0.78 to 2.77) 0.41 (0.35 to 0.47) -0.95 (-4.18 to 2.29)	0.09 1.11 1.28 0.51 0.03 1.65	0.14 0.24 0.13 <0.001 <0.001 0.57
Watt <sub>max</sub> , % predicted Age Sex <i>Pseudomonas aeruginosa</i> Body mass index, <i>z</i> -score FEV <sub>1</sub> , % predicted CFTR group	0.02 (-0.19 to 0.23) 0.23 (-2.21 to 2.66) -2.85 (-5.69 to -0.22) 1.30 (0.19 to 2.42) 0.46 (0.40 to 0.53) -1.38 (-5.04 to 2.27)	0.11 1.24 1.45 0.57 0.03 1.86	0.87 0.85 0.048 0.02 <0.001 0.46

Definition of abbreviations: CFTR = cystic fibrosis transmembrane conductance regulator; CI = confidence interval;  $FEV_1$  = forced expiratory volume in 1 second;  $\dot{V}o_{2peak}$  = peak oxygen consumption, Watt<sub>max</sub> = maximum work load.

The categorical variable *Pseudomonas aeruginosa* is coded as 0 = no and 1 = yes. Sex is coded as 0 for females and 1 for males. CFTR groups are coded as 0 (classes I–III combined) and 1 (classes IV–V combined). Vo<sub>2peak</sub> and Watt<sub>max</sub> percent predicted values were calculated using reference equations from Orenstein (22) and Godfrey and colleagues (23), respectively.

behavior in early childhood leading to a less active lifestyle.

It is important to note that VO<sub>2peak</sub> shows a large variation across the population, and available prediction equations have their limitations. Most of the equations that are used in CF research were established several decades ago, and mostly on the basis of small study samples that did not cover the whole age range of our population and/or did not consider both sexes (21, 22, 36). We decided to use VO<sub>2peak</sub> prediction equations published by Orenstein (22) because these equations were shown to be related to health-related quality of life (37), patient-reported health status (38), and mortality (10) in patients with CF. In addition, we tested another frequently used prediction equation for  $V_{O_{2}peak}$  (21), and the results of all models were comparable to the prediction equation published by Orenstein (22). However, when using equations from Jones and colleagues (21), we noticed significant associations between VO<sub>2peak</sub> (and Watt<sub>max</sub> in percent predicted) with age and sex in the adjusted random effects models, which raises concern about the validity of these equations for this specific CF population.

This study has several limitations. First, we collected data from different

international study centers. Thus, we cannot rule out differences in treatment strategies and treatment quality possibly affecting the health status of the patients, thereby introducing bias. Second, the groups classified according to CFTR classes were unevenly distributed, with a high number of patients in the group with the most common F508del-CFTR mutation (39, 40) compared with patients carrying a CFTR class I, III, IV, or V mutation. Despite the large sample size of 726 patients from 17 different CF centers worldwide, group sizes with CFTR classes I and III-V mutations were relatively small owing to the generally low prevalence of these gene mutations (41). For these reasons, the exploratory analysis comparing exercise capacity among patients with at least one copy of the F508del-CFTR mutation should be interpreted with caution. Moreover, we were not able to consider all known confounders impacting exercise capacity, such as physical activity (9), inflammatory markers, and other CF airway pathogens that were either not routinely assessed or not available for this retrospective analysis. Nevertheless, strong predictors of VO<sub>2peak</sub>, such as pulmonary function (9, 13, 29, 30, 33) and nutritional status (34), as well as proxy measures of inflammation

(i.e., *P. aeruginosa* status), were included in our statistical analysis. In addition, our analysis was limited to maximal CPET outcomes. Additional measures such as  $\dot{V}o_2$ at the anaerobic threshold would have provided further insight into muscle (dys) function. Finally, we acknowledge the limitation of the retrospective study design and the collection of data over a large time period. However, it seems practically impossible to acquire such a large dataset on CPET variables within a prospective study that would overcome these limitations.

In summary, in our large, international cohort of children, adolescents, and adults with CF, we did not detect an association between *CFTR* genotype group and maximal exercise capacity. In this cohort, lower pulmonary function and worse nutritional status were associated with reduced exercise capacity. These findings underline the importance of preserving lung function and maintaining adequate nutrition to prevent exercise intolerance in patients with CF.

**Author disclosures** are available with the text of this article at www.atsjournals.org.

#### Contributors of the CFTR-Exercise Study Group

Donna L. Wilkes, Clinical Research Services, Hospital for Sick Children, Toronto, Ontario, Canada; Greg D. Wells, Division of Respiratory Medicine, Hospital for Sick Children, Toronto, Ontario, Canada; Melinda Solomon, Division of Respiratory Medicine, Hospital for Sick Children, Toronto, Ontario, Canada; Jen Ebidia, Division of Respiratory Medicine, Hospital for Sick Children, Toronto, Ontario, Canada; Stephanie Trgachef, Division of Respiratory Medicine, Hospital for Sick Children, Toronto, Ontario, Canada; Alexandra Woloschuk, Division of Respiratory Medicine, Hospital for Sick Children, Toronto, Ontario, Canada; Esther Quintana-Gallego, Medical-Surgical Unit of Respiratory Diseases, Virgen del Rocio University Hospital, Seville, Spain; Pilar Cejudo, Medical-Surgical Unit of Respiratory Diseases, Virgen del Rocio University Hospital, Seville, Spain; Sibylle Junge, Clinic for Pediatric Pneumology and Neonatology, Hannover Medical School, Hannover, Germany; Christina Smaczny, Medical Clinic I, Pneumology and Allergology, University Hospital Frankfurt/Main, Goethe University, Frankfurt/Main, Germany; Andrew Fall, Department of Respiratory and Sleep Medicine, Royal Hospital for Sick Children, Edinburgh, United Kingdom; Sarah Blacklock, Department of Respiratory and Sleep Medicine, Royal Hospital for Sick Children, Edinburgh, United Kingdom; John Tsanakas, Paediatric Pulmonology and CF Unit, Paediatric Department, Aristotle University of Thessaloniki, Thessaloniki, Greece; Ingrid Kaluza, Department of Pulmonology, Hietzing Hospital, Vienna, Austria.

#### References

- 1 Boyle MP, De Boeck K. A new era in the treatment of cystic fibrosis: correction of the underlying CFTR defect. *Lancet Respir Med* 2013;1: 158–163.
- 2 Cook DP, Rector MV, Bouzek DC, Michalski AS, Gansemer ND, Reznikov LR, *et al.* Cystic fibrosis transmembrane conductance regulator in sarcoplasmic reticulum of airway smooth muscle: implications for airway contractility. *Am J Respir Crit Care Med* 2016; 193:417–426.
- 3 Divangahi M, Balghi H, Danialou G, Comtois AS, Demoule A, Ernest S, et al. Lack of CFTR in skeletal muscle predisposes to muscle wasting and diaphragm muscle pump failure in cystic fibrosis mice. *PLoS Genet* 2009;5:e1000586.
- 4 Lamhonwah AM, Bear CE, Huan LJ, Kim Chiaw P, Ackerley CA, Tein I. Cystic fibrosis transmembrane conductance regulator in human muscle: dysfunction causes abnormal metabolic recovery in exercise. *Ann Neurol* 2010;67:802–808.
- 5 Warth JD, Collier ML, Hart P, Geary Y, Gelband CH, Chapman T, *et al.* CFTR chloride channels in human and simian heart. *Cardiovasc Res* 1996;31:615–624.
- 6 Becq F. CFTR channels and adenosine triphosphate release: the impossible rendez-vous revisited in skeletal muscle. J Physiol 2010; 588:4605–4606.
- 7 Troosters T, Langer D, Vrijsen B, Segers J, Wouters K, Janssens W, et al. Skeletal muscle weakness, exercise tolerance and physical activity in adults with cystic fibrosis. *Eur Respir J* 2009;33:99–106.
- 8 de Meer K, Gulmans VA, van Der Laag J. Peripheral muscle weakness and exercise capacity in children with cystic fibrosis. *Am J Respir Crit Care Med* 1999;159:748–754.
- 9 Hebestreit H, Kieser S, Rüdiger S, Schenk T, Junge S, Hebestreit A, et al. Physical activity is independently related to aerobic capacity in cystic fibrosis. *Eur Respir J* 2006;28:734–739.
- 10 Nixon PA, Orenstein DM, Kelsey SF, Doershuk CF. The prognostic value of exercise testing in patients with cystic fibrosis. N Engl J Med 1992;327:1785–1788.
- 11 Kaplan TA, Moccia-Loos G, Rabin M, McKey RM Jr. Lack of effect of  $\Delta$ F508 mutation on aerobic capacity in patients with cystic fibrosis. *Clin J Sport Med* 1996;6:226–231.
- 12 Selvadurai HC, McKay KO, Blimkie CJ, Cooper PJ, Mellis CM, Van Asperen PP. The relationship between genotype and exercise tolerance in children with cystic fibrosis. *Am J Respir Crit Care Med* 2002;165:762–765.
- 13 van de Weert-van Leeuwen PB, Slieker MG, Hulzebos HJ, Kruitwagen CL, van der Ent CK, Arets HG. Chronic infection and inflammation affect exercise capacity in cystic fibrosis. *Eur Respir J* 2012;39: 893–898.
- 14 McBride MG, Schall JI, Zemel BS, Stallings VA, Ittenbach RF, Paridon SM. Clinical and genetic correlates of exercise performance in young children with cystic fibrosis. *Percept Mot Skills* 2010;110:995–1009.
- 15 Shah AR, Gozal D, Keens TG. Determinants of aerobic and anaerobic exercise performance in cystic fibrosis. Am J Respir Crit Care Med 1998;157:1145–1150.
- 16 McKone EF, Goss CH, Aitken ML. CFTR genotype as a predictor of prognosis in cystic fibrosis. Chest 2006;130:1441–1447.
- 17 Godfrey S, Mearns M. Pulmonary function and response to exercise in cystic fibrosis. *Arch Dis Child* 1971;46:144–151.
- 18 Ballmann M, Rabsch P, von der Hardt H. Long-term follow up of changes in FEV<sub>1</sub> and treatment intensity during *Pseudomonas aeruginosa* colonisation in patients with cystic fibrosis. *Thorax* 1998; 53:732–737.
- 19 Miller MR, Hankinson J, Brusasco V, Burgos F, Casaburi R, Coates A, et al.; ATS/ERS Task Force. Standardisation of spirometry. Eur Respir J 2005;26:319–338.

- 20 Quanjer PH, Stanojevic S, Cole TJ, Baur X, Hall GL, Culver BH, et al.; ERS Global Lung Function Initiative. Multi-ethnic reference values for spirometry for the 3–95-yr age range: the global lung function 2012 equations. *Eur Respir J* 2012;40:1324–1343.
- 21 Jones NL, Makrides L, Hitchcock C, Chypchar T, McCartney N. Normal standards for an incremental progressive cycle ergometer test. *Am Rev Respir Dis* 1985;131:700–708.
- 22 Orenstein DM. Assessment of exercise pulmonary function. In: Rowland TW, editor. Pediatric laboratory exercise testing: clinical guidelines. Champaign, IL: Human Kinetics; 1993. pp. 141–163.
- 23 Godfrey S, Davies CT, Wozniak E, Barnes CA. Cardio-respiratory response to exercise in normal children. *Clin Sci* 1971;40:419–431.
- 24 Fairbarn MS, Blackie SP, McElvaney NG, Wiggs BR, Paré PD, Pardy RL. Prediction of heart rate and oxygen uptake during incremental and maximal exercise in healthy adults. *Chest* 1994;105: 1365–1369.
- 25 de Onis M, Onyango AW, Borghi E, Siyam A, Nishida C, Siekmann J. Development of a WHO growth reference for school-aged children and adolescents. *Bull World Health Organ* 2007;85:660–667.
- 26 Deurenberg P, Weststrate JA, Seidell JC. Body mass index as a measure of body fatness: age- and sex-specific prediction formulas. Br J Nutr 1991;65:105–114.
- 27 Welsh MJ, Smith AE. Molecular mechanisms of CFTR chloride channel dysfunction in cystic fibrosis. *Cell* 1993;73:1251–1254.
- 28 Rowe SM, Miller S, Sorscher EJ. Cystic fibrosis. N Engl J Med 2005; 352:1992–2001.
- 29 Lands LC, Heigenhauser GJF, Jones NL. Analysis of factors limiting maximal exercise performance in cystic fibrosis. *Clin Sci (Lond)* 1992;83:391–397.
- 30 Pastré J, Prévotat A, Tardif C, Langlois C, Duhamel A, Wallaert B. Determinants of exercise capacity in cystic fibrosis patients with mild-to-moderate lung disease. BMC Pulm Med 2014;14:74.
- 31 Hose AJ, Depner M, Illi S, Lau S, Keil T, Wahn U, et al. Latent class analysis reveals clinically relevant atopy phenotypes in 2 birth cohorts. J Allergy Clin Immunol 2017;139:1935–1945.e12.
- 32 Nevill AM. The appropriate use of scaling techniques in exercise physiology. *Pediatr Exerc Sci* 1997;9:295–298.
- 33 Klijn PH, van der Net J, Kimpen JL, Helders PJ, van der Ent CK. Longitudinal determinants of peak aerobic performance in children with cystic fibrosis. *Chest* 2003;124:2215–2219.
- 34 Klijn PH, Oudshoorn A, van der Ent CK, van der Net J, Kimpen JL, Helders PJ. Effects of anaerobic training in children with cystic fibrosis: a randomized controlled study. *Chest* 2004;125: 1299–1305.
- 35 Gruet M, Troosters T, Verges S. Peripheral muscle abnormalities in cystic fibrosis: etiology, clinical implications and response to therapeutic interventions. J Cvst Fibros 2017;16:538–552.
- 36 Hansen JE, Sue DY, Wasserman K. Predicted values for clinical exercise testing. Am Rev Respir Dis 1984;129:S49–S55.
- 37 Hebestreit H, Schmid K, Kieser S, Junge S, Ballmann M, Roth K, et al. Quality of life is associated with physical activity and fitness in cystic fibrosis. BMC Pulm Med 2014;14:26.
- 38 Radtke T, Puhan MA, Hebestreit H, Kriemler S. The 1-min sit-to-stand test—a simple functional capacity test in cystic fibrosis? J Cyst Fibros 2016;15:223–226.
- 39 Cystic Fibrosis Genetic Analysis Consortium. Worldwide survey of the ΔF508 mutation—report from the Cystic Fibrosis Genetic Analysis Consortium. Am J Hum Genet 1990;47:354–359.
- 40 Kerem B, Rommens JM, Buchanan JA, Markiewicz D, Cox TK, Chakravarti A, et al. Identification of the cystic fibrosis gene: genetic analysis. Science 1989;245:1073–1080.
- 41 Bobadilla JL, Macek M Jr, Fine JP, Farrell PM. Cystic fibrosis: a worldwide analysis of CFTR mutations—correlation with incidence data and application to screening. *Hum Mutat* 2002;19:575–606.